Protocol for bead purification

1. Pool each plate of index PCR products in one low-binding tube.

2. Vortex the bead solution and add beads in a 1:1 volume of the PCR products to the PCR pool and invert the tubes a few times. (e.g. 500 ul pooled products = add 500 ul bead solution).

3. Incubate for 5 min on the bench at RT, and thereafter 5-10 min on the magnet track.

4. With the tubes still on the magnet track - remove the supernatant (will not be used later).

5. Wash thoroughly with 500 ul 80% ethanol, leave for 30 sec and remove. Repeat the wash. (still on the magnet)

6. Open the tubes and let dry for 5-10 min (still on the magnet)

7. Remove the tubes from the magnet and dissolve the pellet in 25 ul EB buffer. Leave on the bench for 2 min and thereafter on the magnet for 5 min.

8. Transfer 20 ul to a new low-binding tube. Leave the original tube on the magnet if it is needed to dissolve again.

9. **NOTE:** the 20 ul should be completely clear. Green colour from the polymerase means that the wash has not been thorough enough and the further process will be compromised.

10. Run a gel to confirm the product size (or Bioanalyzer).

11. Measure the DNA concentration on the Nanodrop (ng/ul)

12. Convert the measurements to nM: (X ng/ul /660*750))*1.000.000 and dilute to 4nM

13. Pool 5 ul from each tube in one new tube.